



Ion-Exchangers Product Line(1)



■ **GigaCap Q-650M**

- Highest capacity (BSA; >162 mg/ml)
- High flow rate
- Small & mid MW proteins

■ **SuperQ-650S,M,C**

- Higher capacity (-155mg/ml)
- Oligonucleotide
- small & mid MW proteins

■ **QAE-550C**

- High capacity (-78 mg/ml)
- Stronger retention

■ **DEAE-650S,M,C**

- Capacity; -35mg/ml
- High resolution
- Large MW proteins (Factor VIII)
- Oligonucleotide



■ **GigaCap S-650M**

- Highest capacity(IgG; 136-176 mg/ml)
- High flow rate
- Peptide
- small MW proteins

■ **MegaCap II SP-550EC**

- Higher capacity(Insulin; 95-120mg/ml)
- High flow rate
- Peptide
- small MW proteins

■ **SP-550C**

- High capacity(-120mg/ml)
- Peptide, Small proteins
- Stronger retention

■ **SP-650, CM-650S, M, C**

- Capacity; -60mg/ml
- High resolution
- Large MW proteins



Ion-Exchangers Product Line(2)

- **TSKgel PW bulk resin series**
 - 20 micron, 30 micron
 - Same selectivity to TSKgel PW series analytical column
 - Higher resolution
 - Higher flow rate due to higher mechanical stability of resin than Toyopearl
 - Applicable to 300 L columns
 - Prepacked columns available at 55 mm I.D. x 20 cm and 108 mm I.D. x 20 cm (20 micron)*
 - Prepacked column available at 210 mm I.D. x 30 cm (30 micron)*
 - * ; Special inquiry products
- **TSKgel DEAE-5PW (20), (30)**
 - Sharper peak
 - Better resolution for mid- and larger size proteins
- **TSKgel SuperQ-5PW (20), (30)**
 - Higher capacity than TSKgel DEAE-5PW
 - Applicable to synthetic oligonucleotide, siRNA, etc.
- **SP-5PW**
 - For proteins separation
- **SP-3PW**
 - High capacity and selectivity for insulin
 - For smaller proteins and peptides

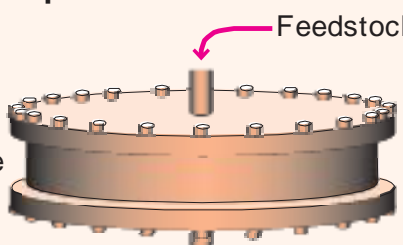
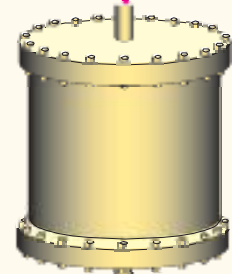
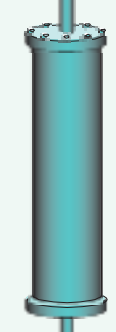

■ Separation of Peptide



Separation of Peptide on Ion-Exchangers

- Purification strategy
- Key factors on separation of peptide
- Cation-exchangers
- Anion-exchangers

Selection of IEC Resins with Different Particle Size

Process step		Bead size	Process media	
			Anion	Cation
Capture		200 μ m		Toyopearl MegaCap SP-550EC
		100 μ m	GigaCap Q-650M Toyopearl SuperQ-650C Toyopearl DEAE-650C Toyopearl QAE-550C	GigaCap S-650M, CM-650M Toyopearl SP-650C Toyopearl SP-550C Toyopearl CM-650C
Intermediate Purification		65 μ m	Toyopearl SuperQ-650M Toyopearl DEAE-650M	Toyopearl SP-650M Toyopearl CM-650M
		35 μ m	Toyopearl SuperQ-650S Toyopearl DEAE-650S	Toyopearl SP-650S Toyopearl CM-650S
Polishing		30 μ m	TSKgel SuperQ-5PW (30) TSKgel DEAE-5PW (30)	TSKgel SP-5PW (30)
		20 μ m	TSKgel SuperQ-5PW (20) TSKgel DEAE-5PW (20)	TSKgel SP-5PW (20)
QC		10 μ m	TSKgel SuperQ-5PW (10) TSKgel DEAE-5PW (10)	TSKgel SP-5PW (10) TSKgel CM-5PW (10)
Pure Product				

Same selectivity HPLC columns are available for most process media



Key Factors on Peptides

- Molecular mass (weight)
 - Applicable to size-exclusion chromatography
- Ionic properties; isoelectric point (pI)
 - pI is determined by total amounts of acidic and basic amino acids
 - Applicable to ion-exchange chromatography
- Hydrophobicity (hydrophilicity)
 - All amino acids contribute to hydrophobicity.
 - Applicable to reversed-phase chromatography
 - Applicable to normal phase chromatography/HILIC
- Metal chelating property (IMAC)
 - His, Met, Tyr, Cys are contribute to interaction.
 - Applicable to metal ion chelating affinity chromatography (IMAC)



Key Factors on Chromatographic Matrix

- Selectivity
 - Functional groups; ionic, hydrophobic, affinity
- Capacity and adsorption strength
 - Amounts and species of functional group
 - e.g. DEAE/QAE, C18/C8
- Particle size
 - 20, 30, 60 100, 200 micron
 - Dependent on purification step, purity (impurity), column volume
- Pore size
 - Larger pore for larger molecule, but causes lower surface.
 - Smaller pore for smaller molecule but causes exclusion from inner surface
- Lot to Lot variation of matrix/resin
 - Selectivity, capacity, recovery, etc.
- Supply of large amounts of resin
 - Scale-up ability and productivity

Key Factors on Chromatography

- Mobile phase/Eluent
 - Buffer species; phosphate, acetate, citrate, Tris,
 - Buffer pH; acidic, basic and neutral
 - Salt as counter ion; NaCl, citrate, acetate, etc.
- Linear or step gradient
 - Higher resolution or easy operation
- Flow rate
 - Productivity
 - Dependent on rigidity of resin/matrix
- Temperature
 - Seasonal temperature change affects to separation.
- **Combination of all factors above decide and contributes to the separation performance.**
 - Loading capacity, resolution, purity, recovery, reproducibility, etc.

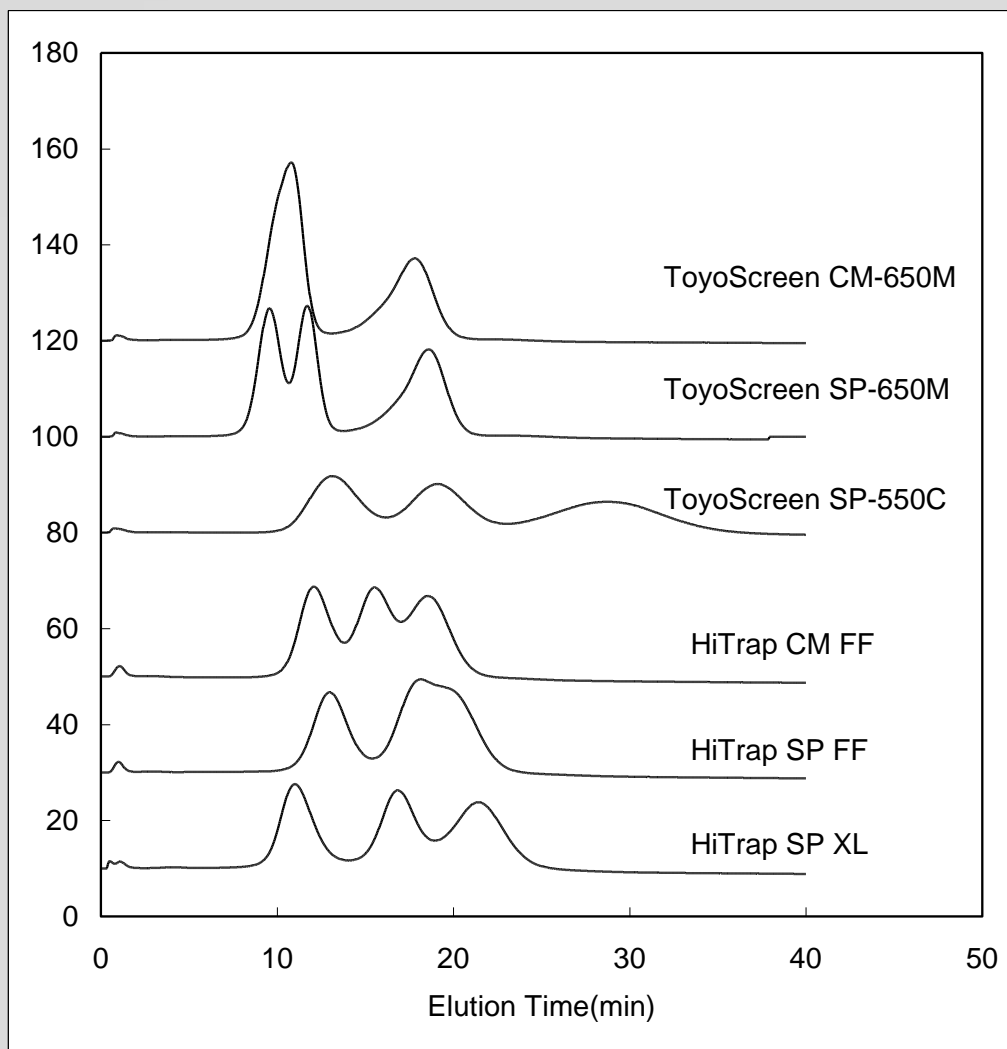
Property of Recombinant Insulin

- MW; ca. 5,800 - 6,100
- Ip; originally ca. 5.5
 - varies around 5.0 – 7.0 by substitution of amino acids
- Impurity for recombinant products
 - Amino acids substitute analogues
 - Dimer
 - Host cell protein

■ Cation-Exchanger



Retention and Selectivity on CIEC for protein Separation



Condition

Column Size: ToyoScreen 6.4 mm I.D.*3cm, 1mL

HiTrap 7mm I.D.*2.5cm, 1mL

Eluent :a) 20mM Phosphate Buffer (pH6.0)

b) 20mM Phosphate Buffer
+ 0.5M NaCl (pH6.0)

Gradient: a) to b) 30min Linear

Flow rate: 1mL/min

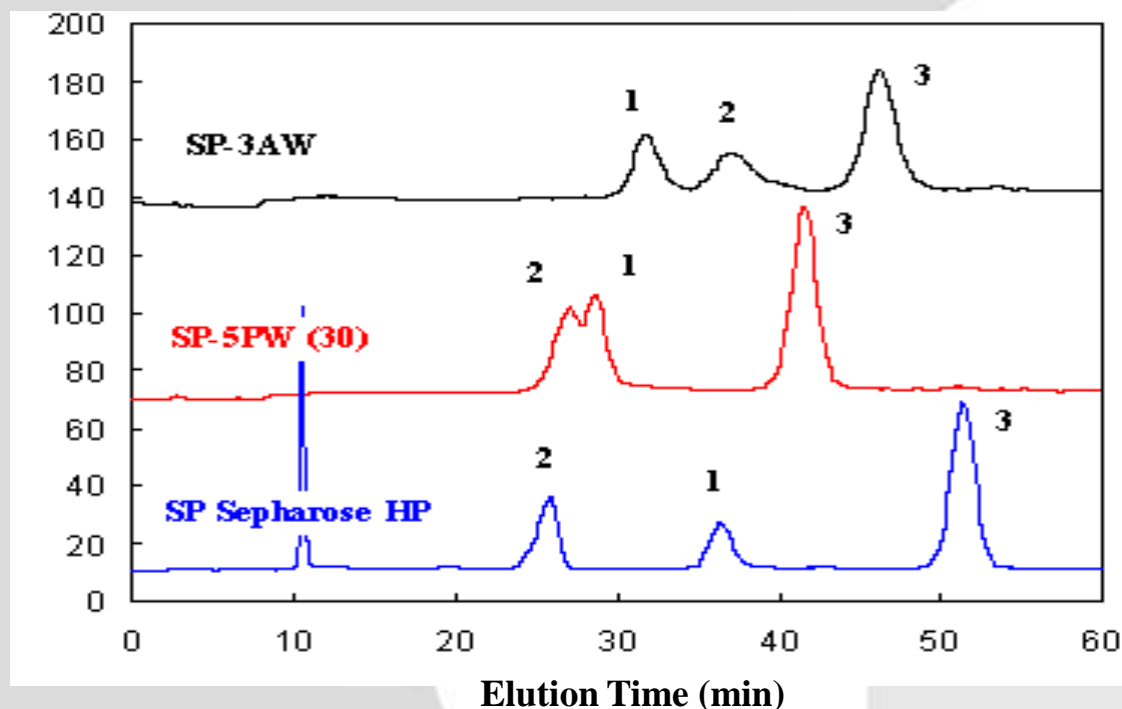
Detection: UV (280nm)

Injection: 50μL

Sample: alpha-Chymotrypsinogen A, Cytochrome C,
Lysozyme 1g/L



Comparison of Elution Behavior of Insulin: SP-3PW(30), SP-5PW(30) and SP Sepharose HP



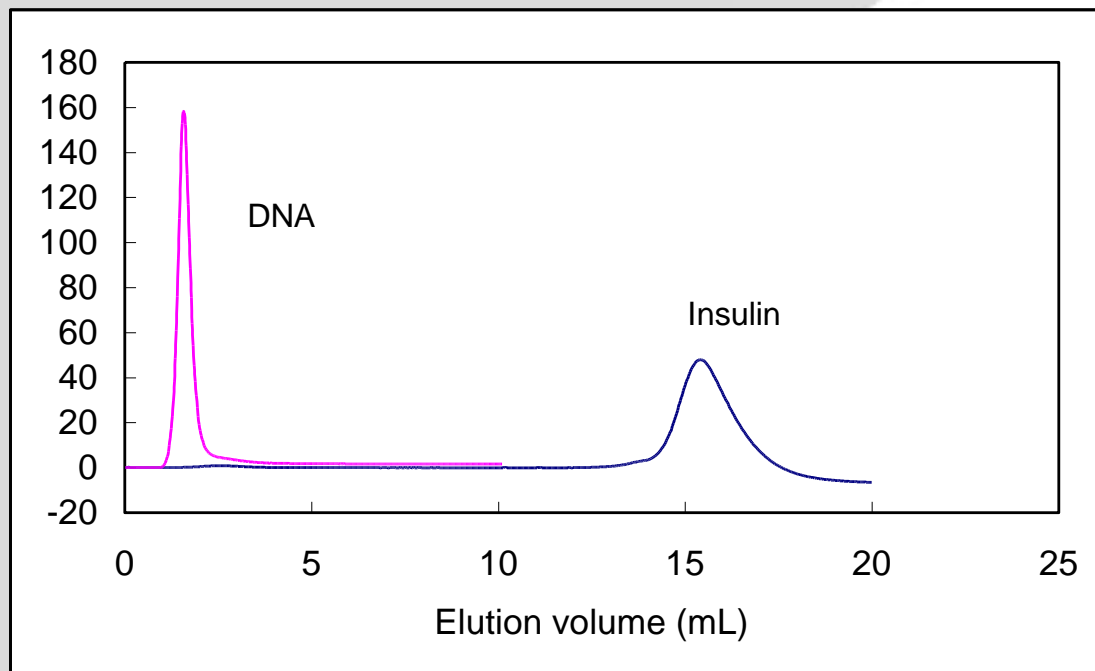
Column Size; 7.5mm I.D. x 7.5cm, Flow Rate; 1.0 ml/min

Sample; 1) Trypsinogen, 2) Insulin, 3) Lysozyme (0.5mg/ml, 100ul injection)

Eluent; A) 20mM Citrate buffer(pH 3.0) / Ethanol = 80:20, B) A + 1.0 M NaCl (pH 3.0) / Ethanol = 80:20
Elution; 60 min. Linear Gradient from A) to B)

TSKgel SP-3PW(30) has stronger retention on insulin.

Separation of Insulin on Toyopearl MegaCap II SP-550EC



Condition

Column: TOYOPEARL MegaCap II SP-550EC(7.5mmID*7.5cm)

Eluent: (A) 0.1M Citrate Buffer (pH3.0)

(B) 0.1M Citrate Buffer (pH6.2)

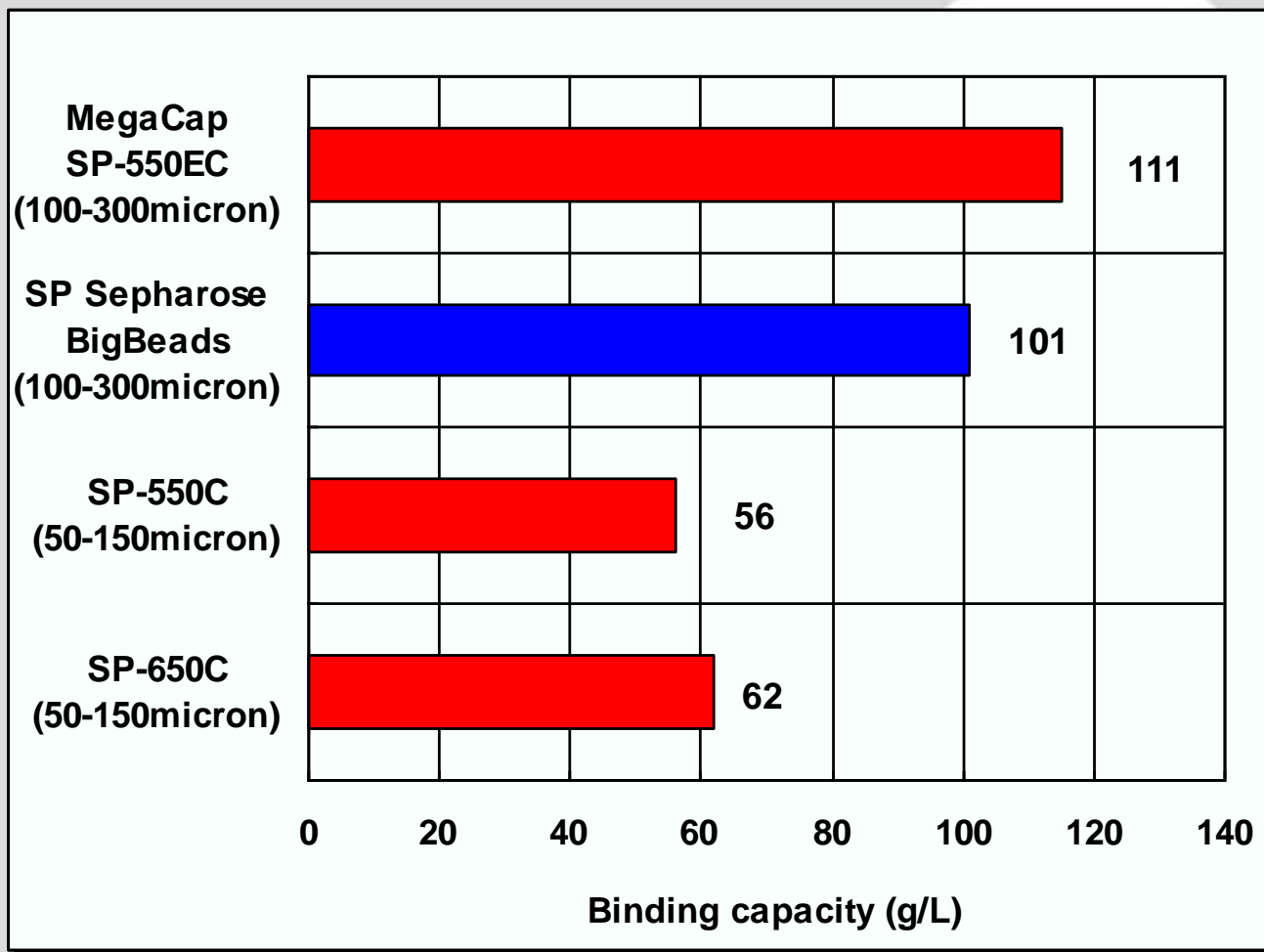
Gradient: (A) --> (B) step at 10min

Flow rate: 1mL/min **Sample:** DNA, Insulin (STD, human) 1g/L

Injection: 100uL **Detection:** UV(280nm)



Comparison of Capacity for Insulin on Cation-Exchangers



* GigaCap S-650M has also higher capacity for insulin.



Comparison of DBC for r-Insulin

	TSKgel SP-3PW (30)	TSKgel SP-5PW (30)	SOURCE 30S
Matrix	polymethacrylate	polymethacrylate	polystyrene divinylbenzen
Particle size	30 micron	30 micron	30 micron
Insulin capacity	49 g/L	24 g/L	45 g/L

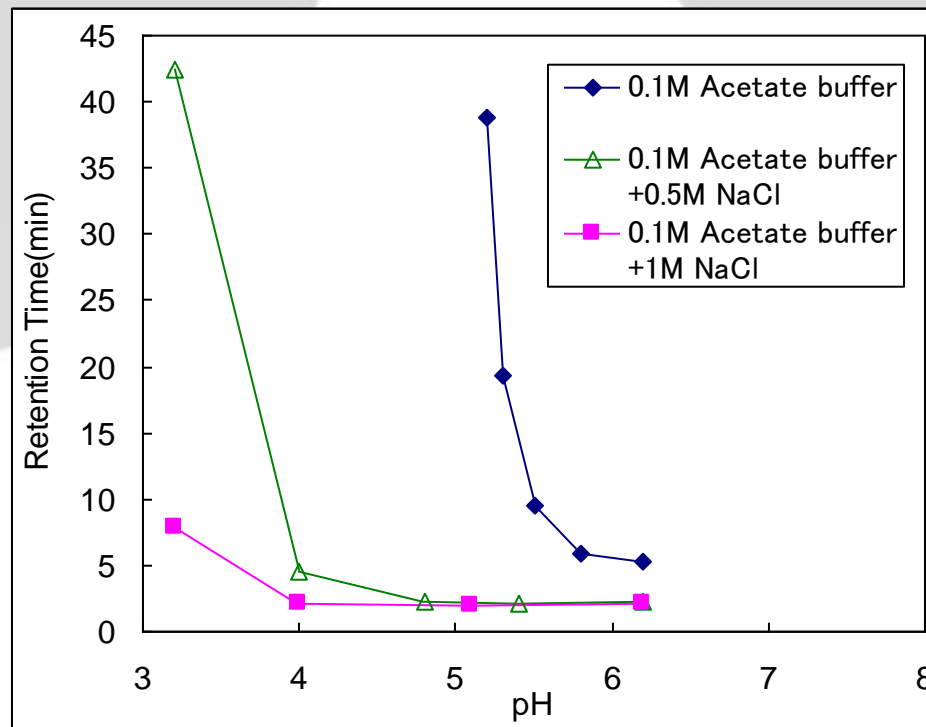
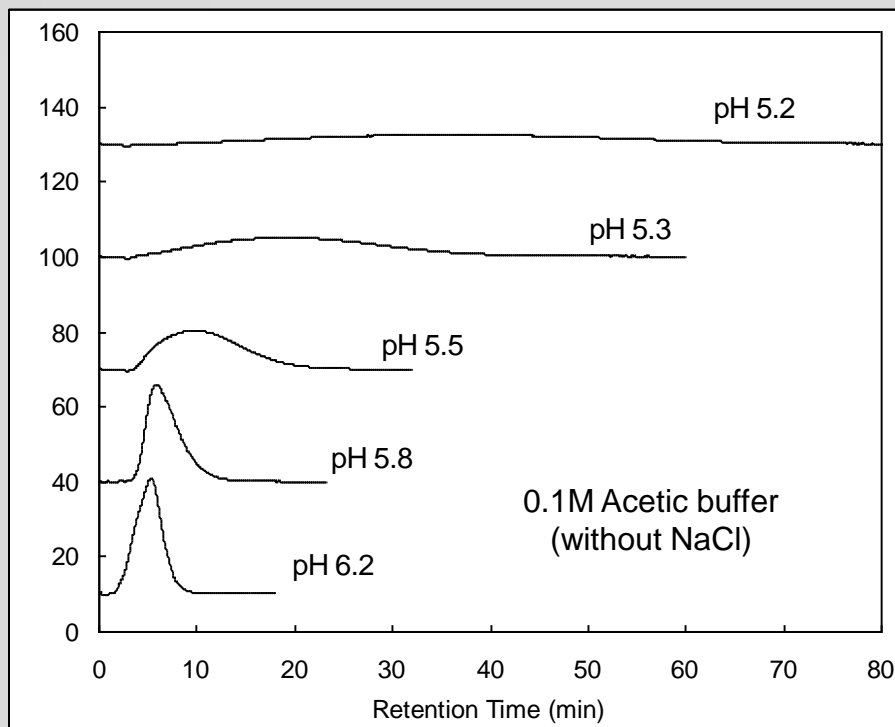
Column; 4.6 mm I.D. x 7.5 cm

Eluent; Gradient elution with 1-propanol by acidic buffer (pH 3.0) containing neutral salt

Flow rate; 0.75 ml/min (270 cm/hr)

Sample; recombinant insulin (7.2 g/L), DBC; calculated by 10 % breakthrough

Effect of pH in Eluent for Insulin Separation by CIEC



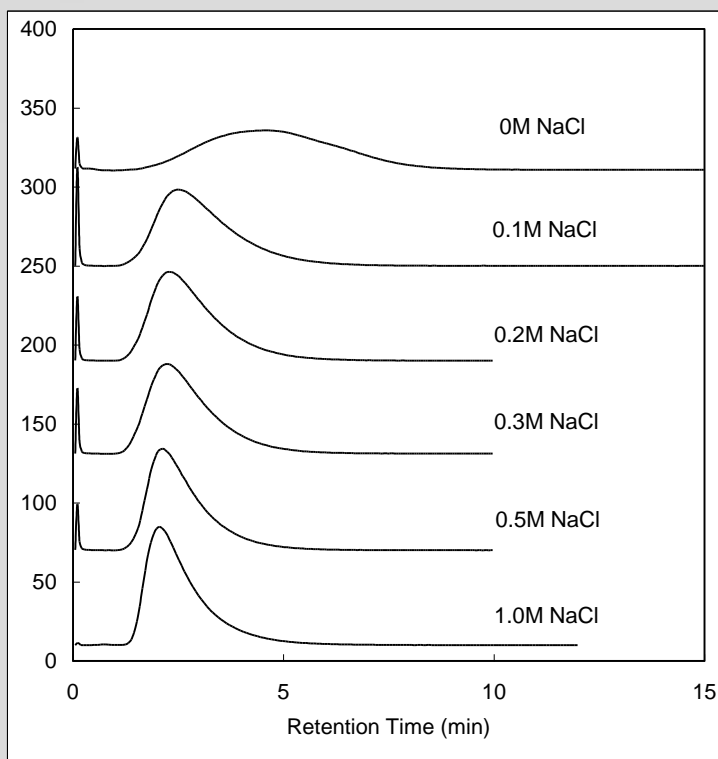
Condition

Column: TOYOPEARL MegaCap II SP-550EC (7.5mmID*7.5cm)

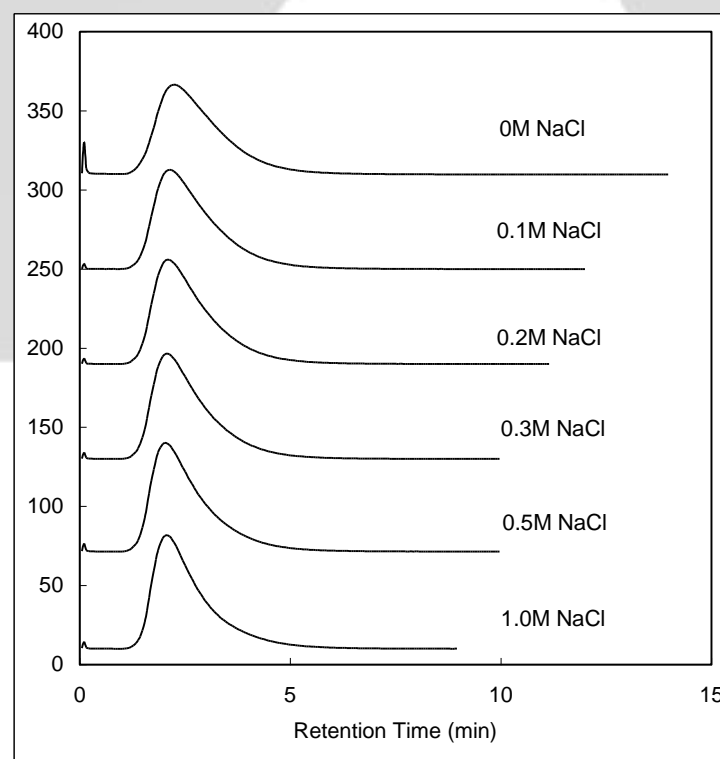
Eluent: 0.1M Aceticate buffer **Flow rate:** 1mL/min **Detection:** UV(280nm)

Sample: Insulin (Human) 1g/L **Injection:** 100uL

Effect of Salt and Buffer for Insulin Separation by CIEC



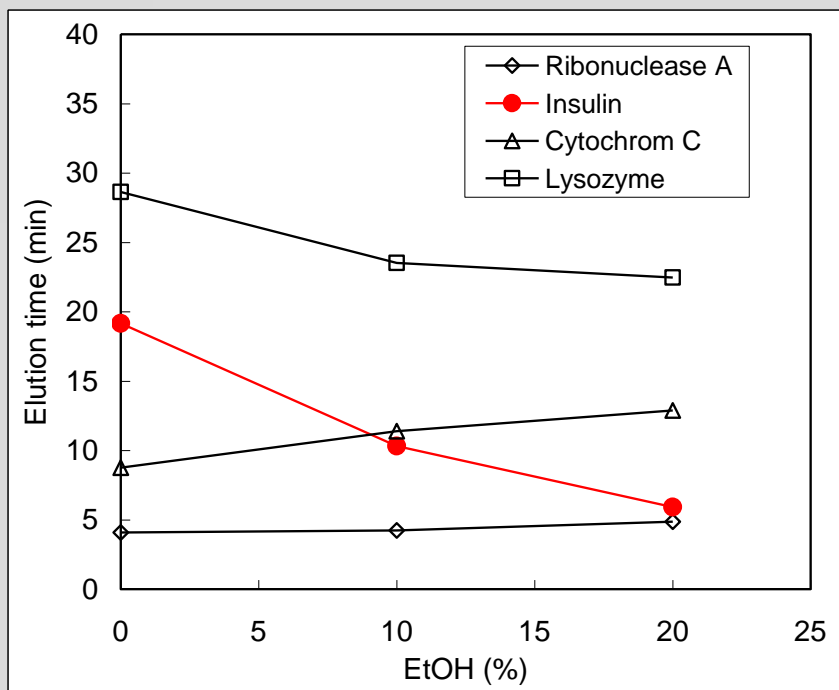
Eluent; Acetate buffer (pH 6.2)



Eluent; Citrate buffer (pH 6.2)

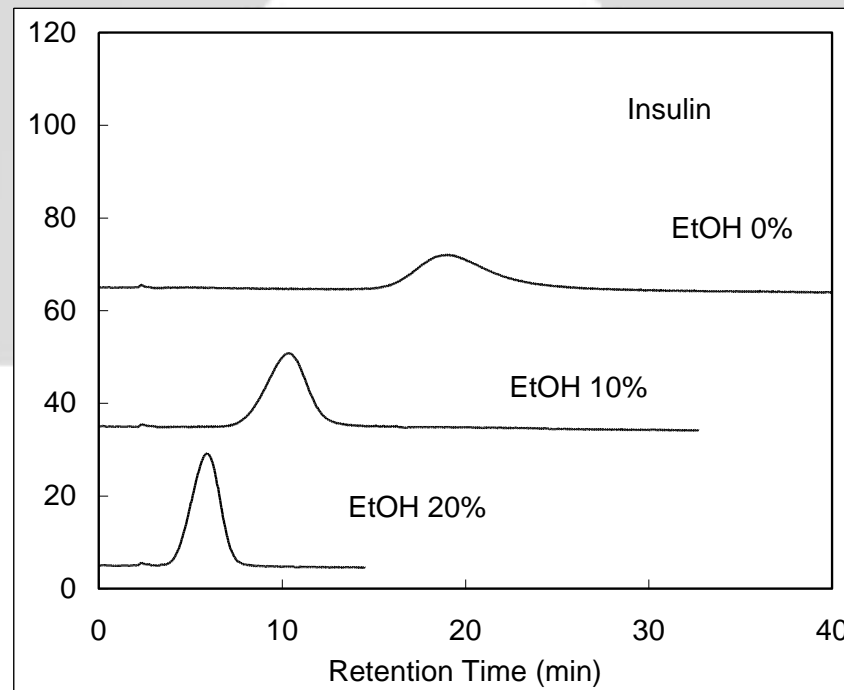
Column; Toyopearl MegaCap II SP-550EC

Effect of Organic solvent for Insulin Separation by CIEC



Condition

Column: IEC-011-LPC (SP resin for Insulin, 20-50um)
 Eluent: (A) EtOH / (0.1M Citrate buffer (pH 5.0))
 (B) EtOH / (0.1M Citrate buffer + 1.0M NaCl (pH5.0))
 Gradient: (A) --> (B) 60min linear
 Flow rate: 1ml/min
 Detection: 280nm(UV)
 Sample: Standard proteins 1g/L
 Injection: 50uL



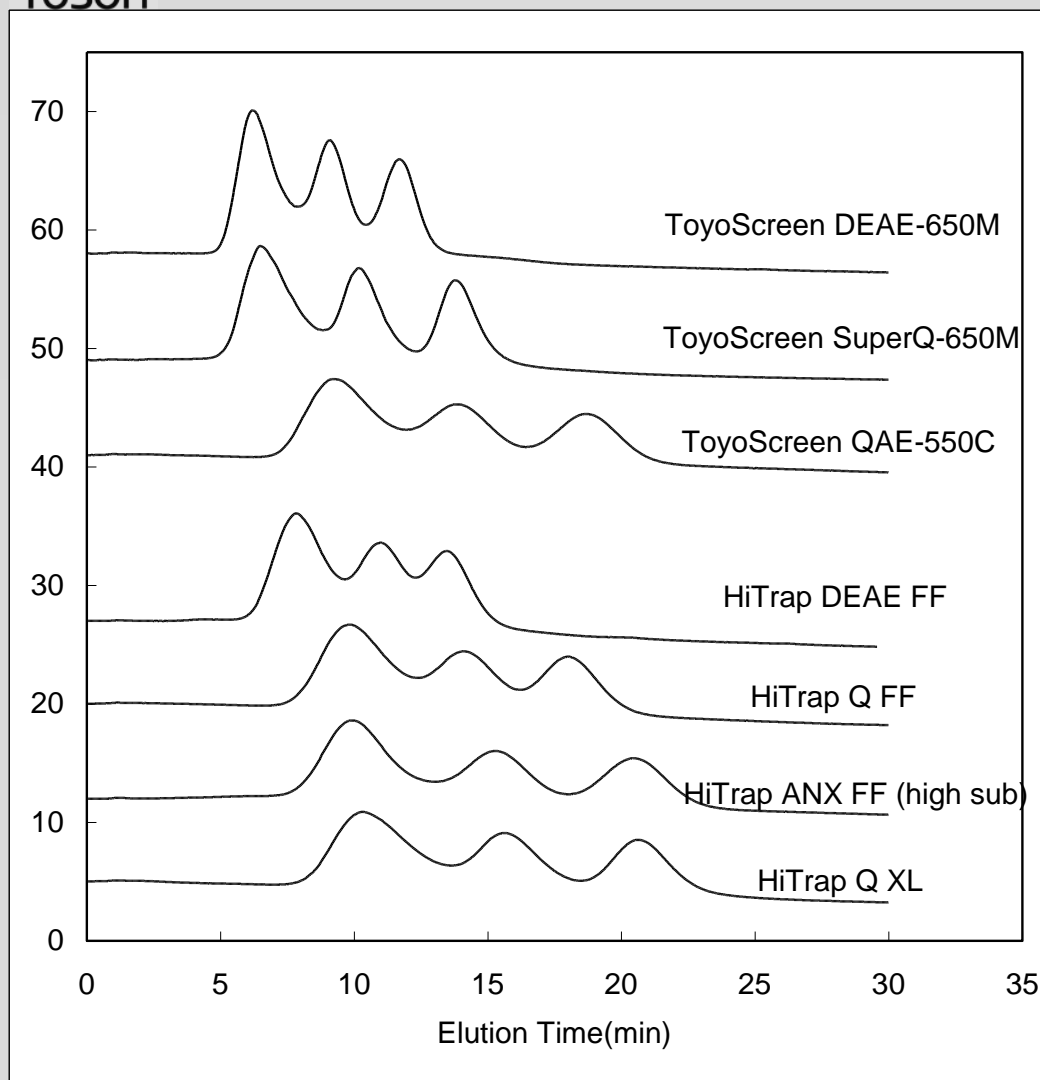
Condition

Column: IEC-011-LPC (SP resin for Insulin, 20-50um)
 Eluent: (A) EtOH / (0.1M Citrate buffer (pH 5.0))
 (B) EtOH / (0.1M Citrate buffer + 1.0M NaCl (pH5.0))
 Gradient: (A) --> (B) 60min linear
 Flow rate: 1ml/min
 Detection: 280nm(UV)
 Sample: Insulin 1g/L
 Injection: 50uL

■ Anion-Exchanger



Retention and Selectivity on AIEC for protein Separation



Condition

Column Size: ToyoScreen 6.4 mm I.D.*3cm, 1mL

HiTrap 7mm I.D.*2.5cm, 1mL

Eluent :a) 20mM Tris-HCl (pH8.0)

b) 20mM Tris-HCl + 0.5M NaCl (pH8.0)

Gradient: a) to b) 30min Linear

Flow rate: 1mL/min

Detection: UV (280nm)

Injection: 50μL

Sample: Transferrin, Ovalbumin,
Trypsin inhibitor 1g/L



Dynamic Binding Capacity of Anion-Exchangers for Insulin

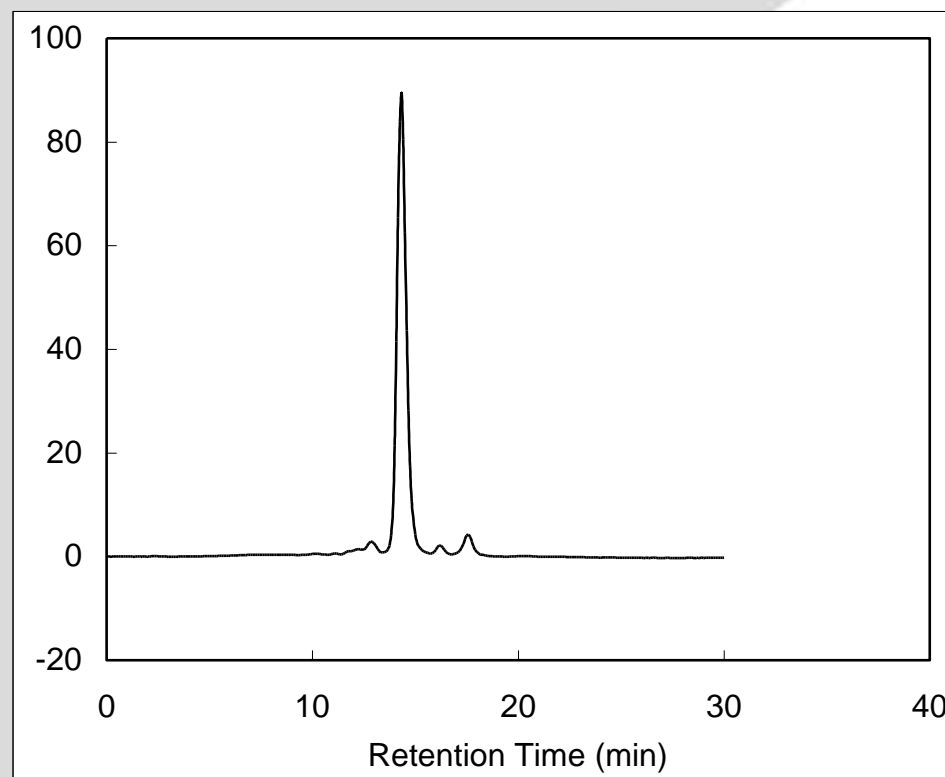
Binding Capacity(g/L-gel) (10% Leakage)	
DEAE-650M	29
QAE-550C	83
SuperQ-650M	162
DEAE Sepharose FF	66



Comparison of Capacity for Insulin on Anion-Exchangers

	TOYOPEARL SuperQ-650M	DEAE Sepharose FF
Base material	Hydrophilic polymer	Cross linked agarose
Particle size	40-90 um	45-165 um
Ion exchange capacity	0.20-0.30 eq/L-gel	0.11-0.16 eq/L-gel
Binding capacity for Insulin	162 g/L-gel	66 g/L-gel
Rigidity	+++	+
Productivity	+++	+

Separation of recombinant Insulin by IEC on TSKgel SuperQ-5PW Analytical Column



Condition

Column: TSKgel SuperQ-5PW (7.5mmID x 7.5cm)

Eluent: (A) EtOH/10mM Tris-HCl (pH 8.7)=1/1

(B) 0.2M NaCl in (A)

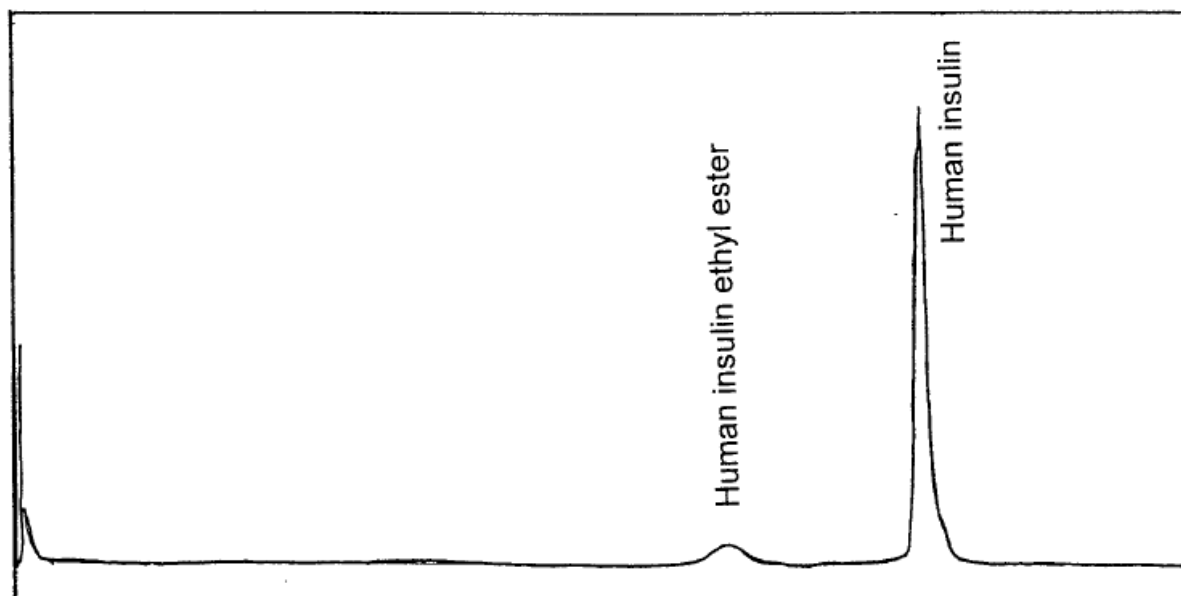
Gradient: (A) --> (B) 30min linear

Flow rate: 1ml/min

Detection: 280nm(UV)

Injection: 50uL

Separation of Insulin and DES-B30-Insulin by IEC on TSKgel SuperQ-5PW



Chromatogram of Example 13

Column; TSK-Gel Q-5PW (Tosoh) column (20 mL)

Elution; Buffer (A); 0.15% w/w triethanolamine, 42.5% w/w ethanol, pH 7.5.

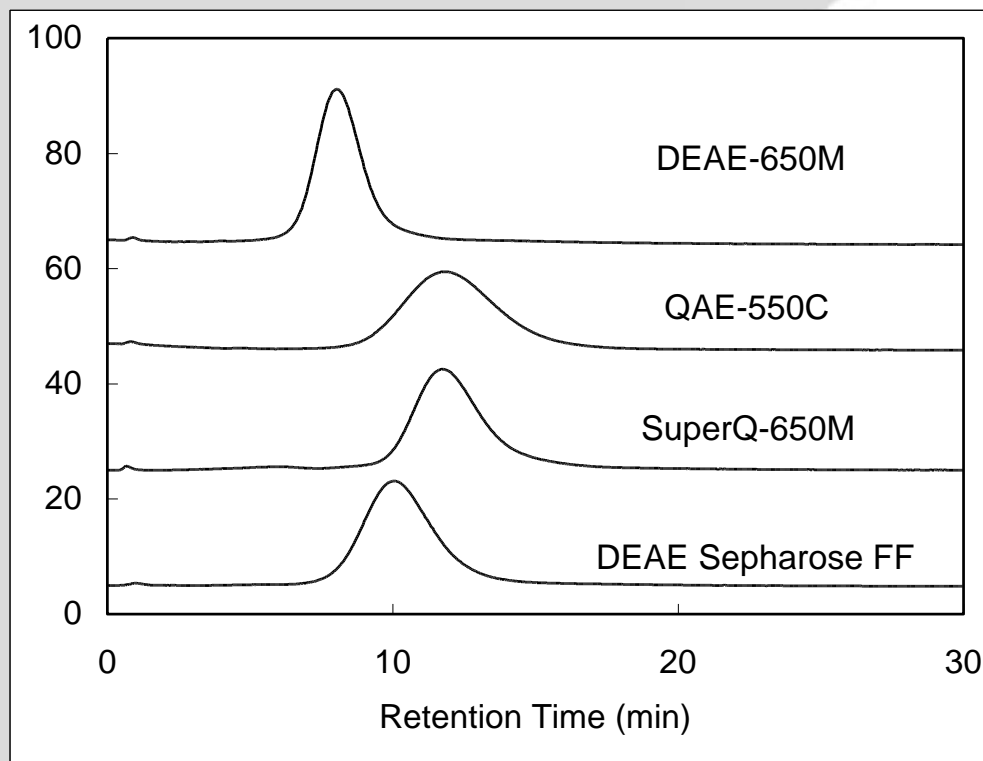
a linear gradient from 0 to 1.14% w/w sodium citrate tri-hydrate in Buffer (A)

Sample; A mixture of 7.7 mg/ml human insulin and 0.8 mg/ml human insulin ethyl ester (B30)

The mixture (2 mL) contained 4 mmol/l EDTA, 16% w/w ethanol, pH 7.5.

Ref. EU patent WO0055184

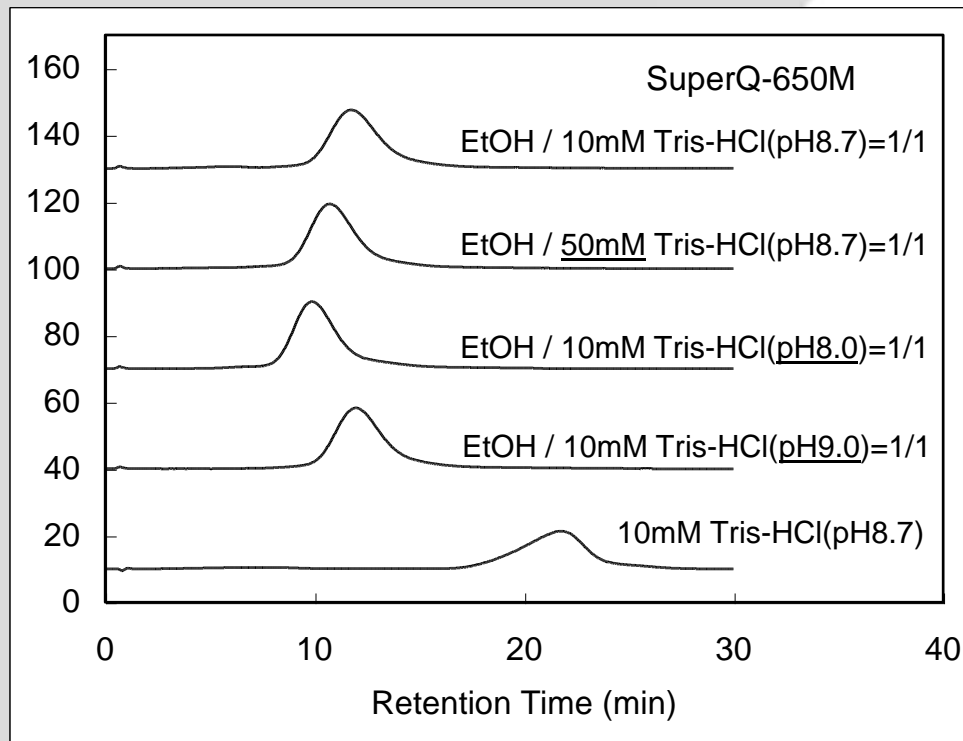
Retention and Selectivity of Insulin on AIEC resins



Condition

Column: ToyoScreen (1mL)
 Eluent: (A) EtOH/10mM Tris-HCl (pH 8.7)=1/1
 (B) 0.2M NaCl in (A)
 Gradient: (A) --> (B) 30min linear
 Flow rate: 1ml/min
 Detection: 280nm(UV)
 Injection: 50uL

Effect of pH in Eluent for Insulin Separation by AIEC



Condition

Column: ToyoScreen SuperQ-650M (1mL)

Eluent: (A) shown in figure

(B) 0.2M NaCl in (A)

Gradient: (A) --> (B) 30min linear

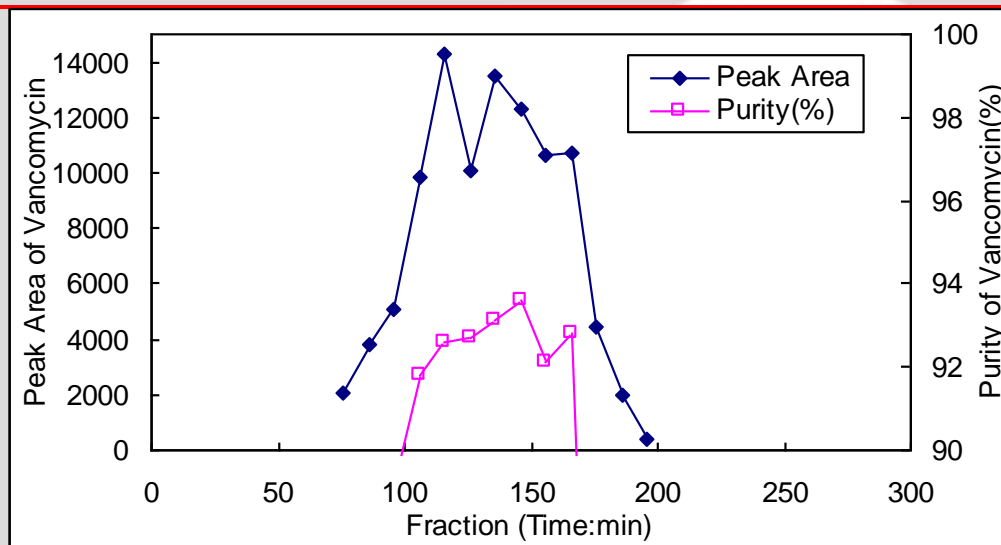
Flow rate: 1ml/min

Detection: 280nm(UV)

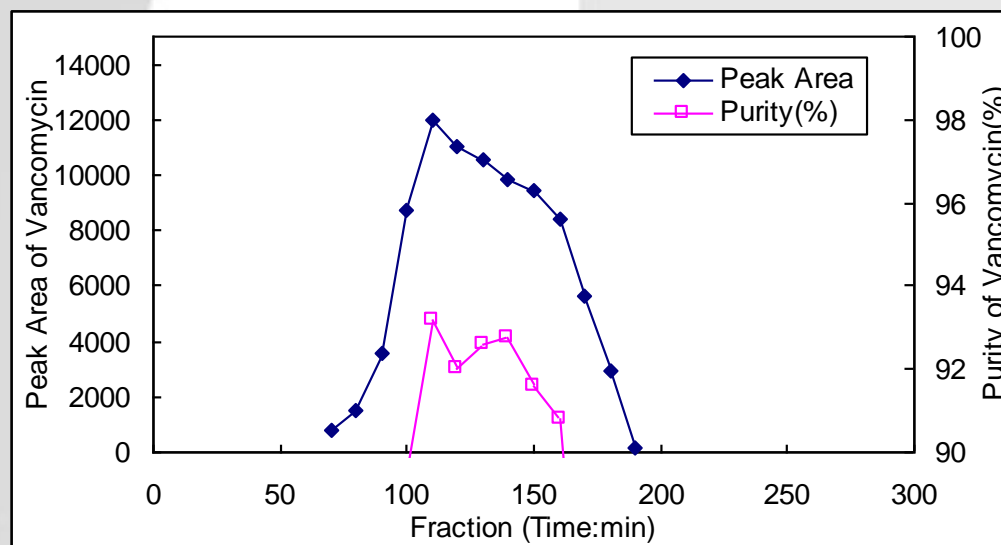
Injection: 50uL

Effect of Temperature on Separation and Purity of Peptide by AIEC

25 C



32 C





Reversed-phase Chromatography

- Selection of stationary phase
 - Monomeric vs. polymeric
 - ODS vs. other functional group C8, C4
 - Endcapping of silanol groups
 - Pore size; 8 – 12 nm
- Selection of Particle size
 - 20-100 micron
- Mobile phase
 - Organic modifier; methanol, acetonitrile
 - Acidic eluent; THF, HCl, acetic acid
 - Gradient method; linear or step
- Sample loading
 - Overloading conditions
- Column cleaning
 - CIP with NaOH for polymer resin
- Process economic
 - Purity, recovery, production efficiency

Process R&D - TOYOSCREEN

- Easy to select the most effective resin for purification
- Selectivity, purity, recovery, capacity and elution conditions
- A-IEC, C-IEC, HIC; Kit and individual grade including AFC
- 1 ml and 5 ml column
- Installation to HPLC and/or Bio-LC system (e.g. AKTA Prime)

